# Collection and Dissection of *Drosophila* pupal genitalia

## Reagents

Nuclease Free H20 GE SH30538.02

Ethanol 200 proof Decon Laboratories #2716

Methanol Fisher A412P-4

16% Paraformaldehyde (PFA) EMS #15710

### **Collect Prepupae**

Search vials for white prepupae and move to a clean glass viewing dish.

Sort males and females to separate wells by looking for larval testis, two clear circles on the ventral/lateral sides of the pupa, which are only present in males.

Place all samples you want to dissect later on a Kimwipe in a petri dish and incubate at 25°C before dissection.

# Dissection of pupal genitalia

Fill a micro-centrifuge tube at least half way with 4% paraformaldehyde in PBT, and place on ice.

Move pupae from incubator to a clean glass viewing well.

Fill 3 other glass viewing wells with PBS.

Impale a pupa in the anterior half with the a microforcep and move to a well with PBS. Using spring scissors to cut the bottom 2/5ths of the pupae off. Discard anterior 3/5s of the pupae.

Move the posterior tip of the pupa to a clean well.

Using a 200µL tip gently flush most of the fat bodies, but try not to wash out the pupal genitalia. Some lab members wash out the genitalia at this step, but it can be difficult to locate the sample amid all the fat bodies, especially after several dissections.

Transfer pupal genitalia to a clean well with PBS and gently wash the sample until you flush the pupal genitalia from the sample.

Transfer the pupal genitalia to the third clean well. *This serves as a final wash before fixation*.

Using a  $20\mu L$  pipet transfer the sample in ~2ul of PBS to the 4% PFA solution and place on ice. We collect samples in 4% PFA on ice, but we do not consider fixation to begin until the sample is placed at room temperature.

Repeat previous steps until all pupae are dissected or 1 hour has past.

#### Fix samples

Incubate samples in 4% PFA at room temperature for 30 minutes.

Wash 2X quickly with methanol.

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Store samples in ethanol at -20°C indefinitely.